# Application of Population Pharmacokinetic Analysis for Quantification of in vivo Binding Properties in the Rat Brain by Positron Emission Tomography 

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## Introduction

The $G A B A_{A}$-receptor plays a significant role in epileptogenesis. For the characterisation of the binding properties of the GABA ${ }_{A}$ receptor in vivo, Positron Emission Tomography (PET) with ${ }^{11} \mathrm{C}$-labelled flumazenil (FMZ) is an attractive technique. In the presented approach a single saturating dose of $\left[{ }^{11} \mathrm{C}\right] F M Z$ is administered and the time courses of the concentration of FMZ in blood (using HPLC-UV) and brain (using PET) are determined.

## Experimental Design

Different dosages of ${ }^{11} \mathrm{C}$-FMZ were tested: $2000 \mu \mathrm{~g}(\mathrm{n}=2), 1000 \mu \mathrm{~g}(\mathrm{n}=1), 500 \mu \mathrm{~g}(\mathrm{n}=7)$, $100 \mu \mathrm{~g}(\mathrm{n}=3), 50 \mu \mathrm{~g}(\mathrm{n}=3), 25 \mu \mathrm{~g}(\mathrm{n}=2)$, and $1 \mu \mathrm{~g}(\mathrm{n}=6)$.

The bloodsamples of the experiments with a dose of $1 \mu \mathrm{~g}$ could not be analysed because the concentrations were below the detection limit of the HPLC-UV system ( $25 \mathrm{ng} / \mathrm{ml}$ ).

## PK-model

A user defined model (figure 2) was implemented in NONMEM's ADVAN9 subroutine. The parameters were optimised using the FO-method.


Figure 2: User defined structural PK-model.

## Results \& Discussion

- The PK-model with saturable binding in the brain is indeed able to describe the observed dose dependent PK of FMZ in blood with sharper peaks at higher dose levels (figure 3).

Model predictions at higher concentrations in the brain are slightly biased (figure 3 and figure 4), which might be explained by FMZ present in blood in the brain. This could not be taken into account with the current dataset.

Including interindividual variability in CL and $\mathrm{V}_{\mathrm{B}}$ resulted in better diagnostics, however, the estimation of different parameters, including $\mathrm{B}_{\max }$ and $\mathrm{K}_{\mathrm{D}}$, was less accurate (\%CV up to $400 \%$ ).

All parameters, except $K_{D}$ are accurately estimated (table 1, model A). While $V_{B r}$ and $Q_{B r}$ have similar values, in the subsequent model $B V_{B r}$ was set to $Q_{B r}$, which resulted in precise estimates of all parameters (table 1).

The parameter estimates are similar to literature values: CL=19.1 vs 33 $\mathrm{ml} / \mathrm{min}, \mathrm{V}_{\mathrm{d}, \mathrm{ss}}=243$ vs 299 ml respectively ${ }^{1}$. $\mathrm{K}_{\mathrm{D} \text { 'in vitro }}{ }^{2}$ and $\mathrm{K}_{\mathrm{D}, \text { in vivo }}$ as presently estimated did not significantly differ ( $7.1 \mathrm{ng} / \mathrm{ml}$ and $4.5 \mathrm{ng} / \mathrm{ml}$ respectively).

Table 1: Population estimates of parameters with coefficient of variation (\%) between brackets.

| parameter | $\mathrm{V}_{\mathrm{C}}$ <br> $(\mathrm{ml})$ | CL <br> $(\mathrm{ml} / \mathrm{min})$ | $\mathrm{V}_{\mathrm{T}}$ <br> $(\mathrm{ml})$ | Q <br> $(\mathrm{ml} / \mathrm{min})$ | $\mathrm{V}_{\mathrm{Br}}$ <br> $(\mathrm{ml})$ | $\mathrm{Q}_{\mathrm{Br}}$ <br> $(\mathrm{ml} / \mathrm{min})$ | $\mathrm{B}_{\max }$ <br> $(\mathrm{ng} / \mathrm{ml})$ | $\mathrm{K}_{\mathrm{D}}$ <br> $(\mathrm{ng} / \mathrm{ml})$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| model A | 38.9 | 18.9 | 179 | 20.3 | 25.4 | 26.3 | 14.1 | 4.6 |
|  | $(23.3)$ | $(4.0)$ | $(15.3)$ | $(15.7)$ | $(16.9)$ | $(21.7)$ | $(39.1)$ | $(50.8)$ |
| model B | 41.9 | 19.1 | 178 | 21.4 | 23.4 | 13.8 | 4.5 |  |
|  | $(17.9)$ | $(3.3)$ | $(14.9)$ | $(8.7)$ | $(21.8)$ | $(22.0)$ | $(18.4)$ |  |
| model A: all parameters estimated |  |  |  |  |  |  |  |  |

model B: $V_{B r}=Q_{B r}$

## Conclusions

- With the presented full saturation approach, using PET and population analysis, PK of FMZ in blood and brain, including binding to the cerebral GABA $A_{A}$ receptor are determined in vivo.
- This method allows simultaneous, independent, and precise estimation of $B_{\max }$ and $K_{D}$ in contrast to reported approaches.

The concentration-time curves of FMZ in blood showed dose dependent PK. It is hypothesised that this can be explained by a considerable amount of FMZ bound to the $\mathrm{GABA}_{\mathrm{A}}$ receptor in the brain. Therefore, the objective was to develop a model, which simultaneously describe the concentrations in blood and brain, and identify binding properties of the $\mathrm{GABA}_{\mathrm{A}}$ receptor in vivo.


Figure 1: Experimental design.

| Blood | $\frac{d A B}{d t}=\operatorname{Rinf}-\mathrm{k} 12 \cdot \mathrm{AB}+\mathrm{k} 21 \cdot \mathrm{AT}-\mathrm{k} 10 \cdot \mathrm{AB}-\mathrm{k} 13 \cdot \mathrm{AB}+\mathrm{k} 31 \cdot \mathrm{CBrF} \cdot \mathrm{VBr}$ |
| :--- | :--- |
| Tissue | $\frac{\mathrm{dAT}}{\mathrm{dt}}=\mathrm{k} 12 \cdot \mathrm{AB}-\mathrm{k} 21 \cdot \mathrm{AT}$ |
| Free in Brain | $\frac{\mathrm{dCBrF}}{\mathrm{dt}}=\mathrm{k} 13 \cdot \frac{\mathrm{AB}}{\mathrm{VBr}}-\mathrm{k} 31 \cdot \mathrm{CBrF}-\mathrm{kon} \cdot(\mathrm{Bmax}-\mathrm{CBrB}) \cdot \mathrm{CBrF}+\mathrm{koff} \cdot \mathrm{CBrB}$ |
| Spec. bound <br> in Brain | $\frac{\mathrm{dCBrB}}{\mathrm{dt}}=\mathrm{kon} \cdot(\mathrm{Bmax}-\mathrm{CBrB}) \cdot \mathrm{CBrF}-\mathrm{koff} \cdot \mathrm{CBrB}$ |
| $\mathrm{K}_{\mathrm{D}}$ | $\mathrm{KD}=\frac{\mathrm{koff}}{\mathrm{kon}}$ |

The residual error was assumed to be proportional to the concentration in blood and brain. Furthermore, to account for the greater uncertainty in blood concentrations that are close to the detection limit an extra residual error was added. This residual variance was fixed to the square of half of the detection limit.


Figure 3: Concentration-time profiles for FMZ in blood and brain after different iv doses (1-2000 $\mu \mathrm{g}$ ) to male Wistar rats. The dots represent the observed FMZ concentrations and the lines represent the individual predictions of model B.


Figure 4: Diagnostic plots for FMZ concentrations in blood (red) and brain (blue).

In future, studies in animal models of epilepsy will be performed using the presented approach, to investigate whether changes in $G A B A_{A}$ receptor properties contribute to the mechanisms of epileptogenesis and pharmacoresistance.

## References:

Mandema et al. (1991) J. Pharmacol. Exp. Ther. 257:472-47

