

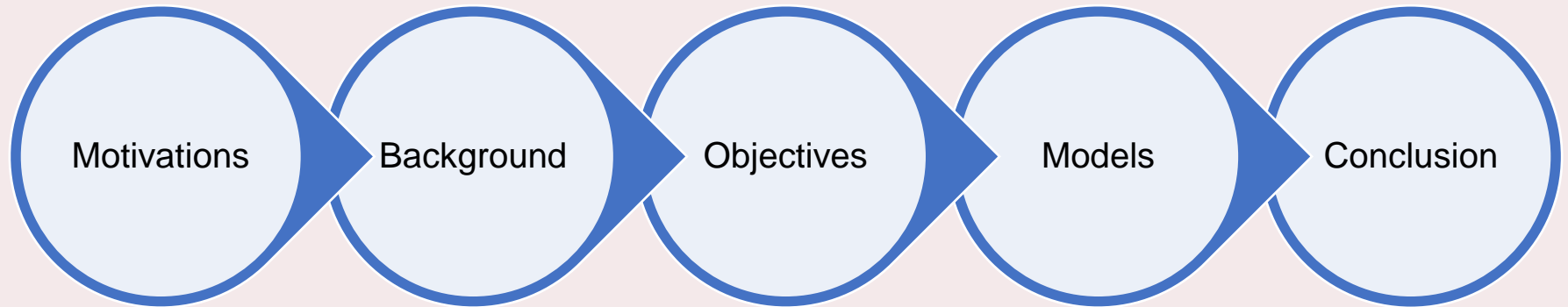
Mathematical Modelling of CD19⁺ B Cell Reconstitution in Children After Insult to the Immune System

Soumya Perinparajah*, Reem Elfeky, Juliana M.F. Silva, John Booth, Oscar J. Charles, S.Y. Amy Cheung, James W.T. Yates, Austen Worth, Judith Breuer, Nigel Klein, Persis J. Amrolia, Paul Veys, Joseph F. Standing

*PhD candidate, Infection Section

Infection, Immunity and Inflammation Research and Training Department
Great Ormond Street Institute of Child Health, University College London, UK







Previous immune reconstitution studies following haematopoietic stem cell transplantation (**HSCT**)



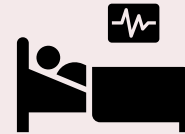
- T cells
- Adults
- Survival analysis

Unmet Need

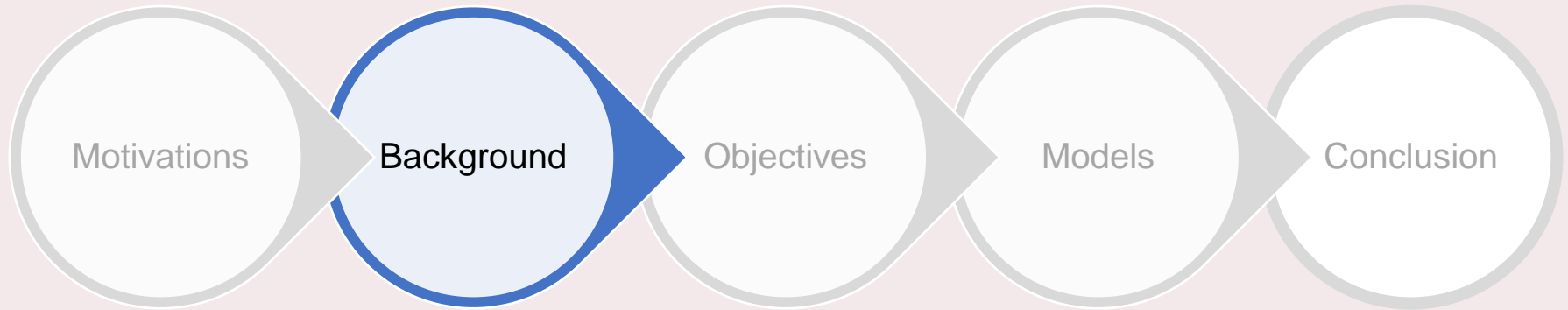


- B cells
- Children
- Pharmacometrics

Clinical Impact

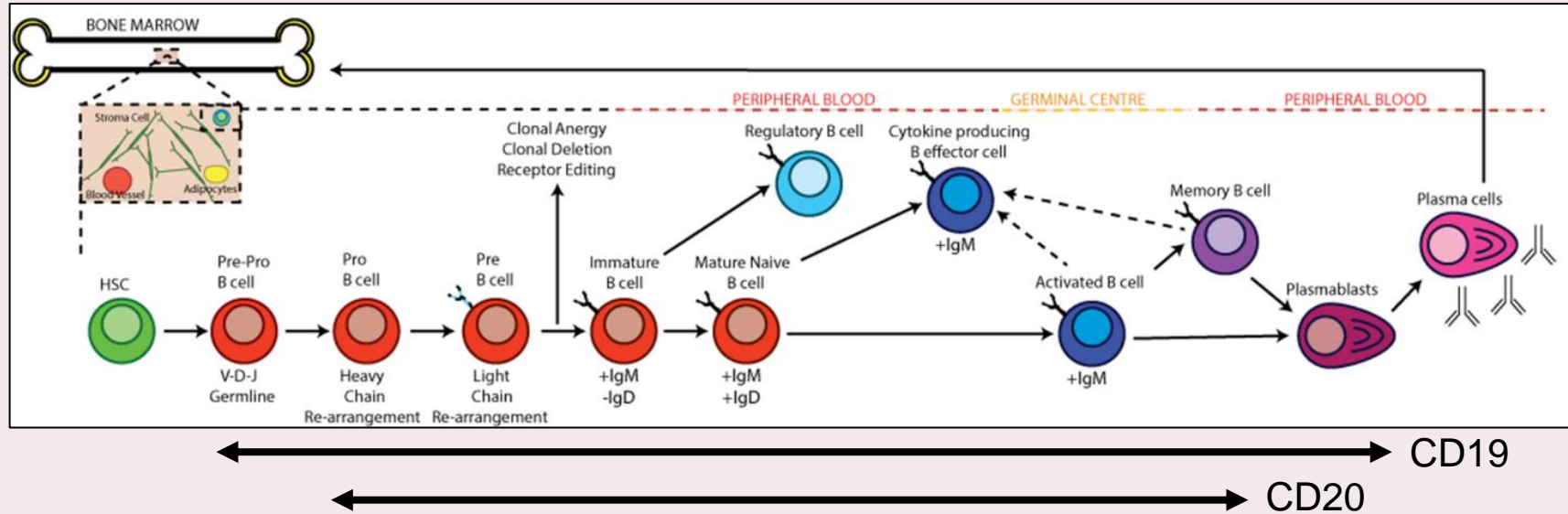


- Key factors associated with B cell recovery
- Treatment decisions
- Predict B cell recovery trajectories



Background

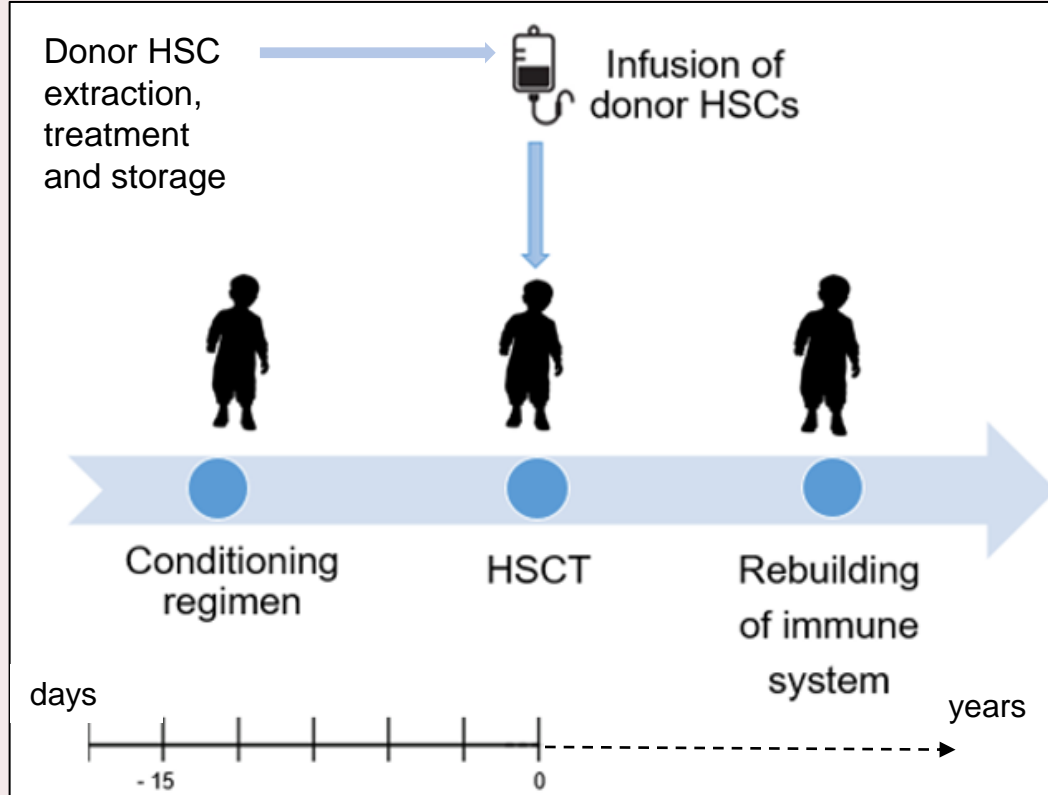
B cells: key function in adaptive immunity as antibody-producing and antigen-presenting cells



Wilkinson and Rosser (2019) *Front. Immunol* [1]

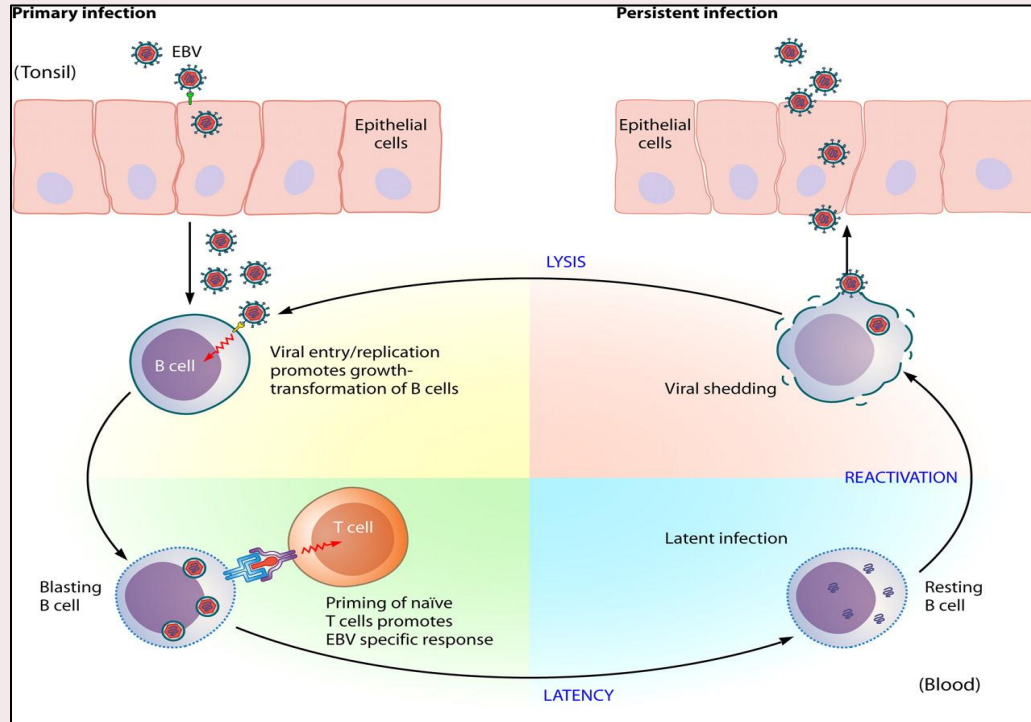
Background

HSCT : Procedure to replace damaged HSCs with healthy HSCs



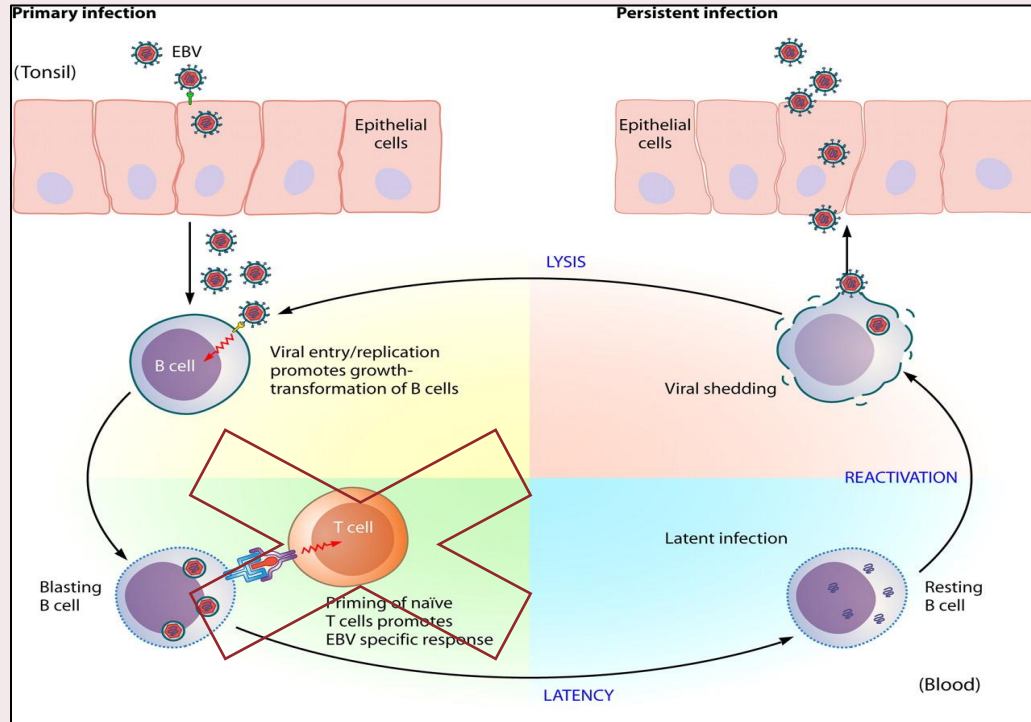
Background

Epstein-Barr Virus (EBV): Commonly **reactivated** post-HSCT due to reduced EBV-specific cytotoxic T cells



Background

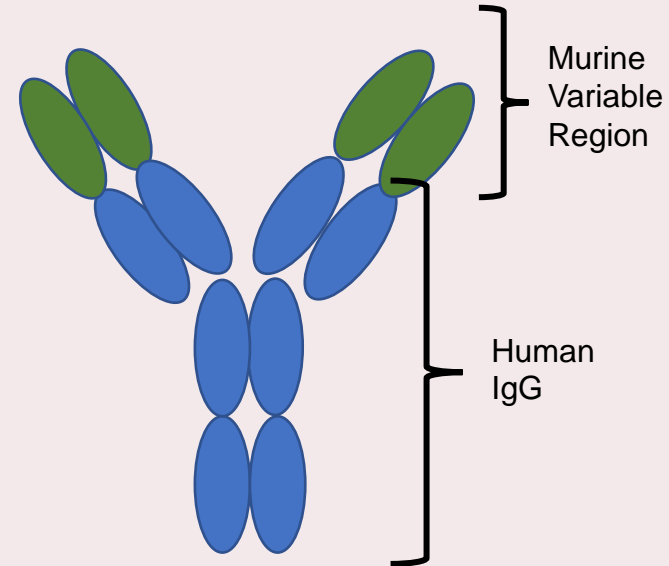
EBV: EBV reactivation is the leading cause of post-transplant lymphoproliferative disease (**EBV-PTLD**)

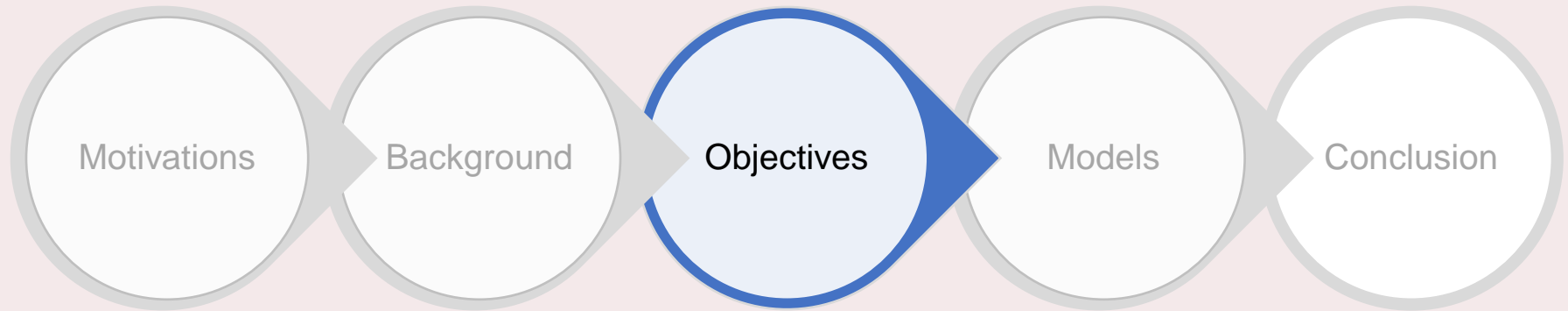


Background

Rituximab: Prescribed off-label for children with EBV post-HSCT

- Chimeric **monoclonal antibody** targeting **CD20⁺** on B cells
- **Limited licensure in children**
 - Rare vascular disorders
 - Previously untreated B lymphoma
- Reported half-life
 - **Adults**
 - 17.4 days (*Gibiasky et al (2021) CPT:PSP [3]*)
 - 19.7 days (*Ng et al (2005) J Clin Pharmacol [4]*)
 - **Children**
 - 26-29 days (*Barth et al (2013) BJH [5]*)
 - 19.3 days (*Pan et al (2019) BJCP [6]*)





1

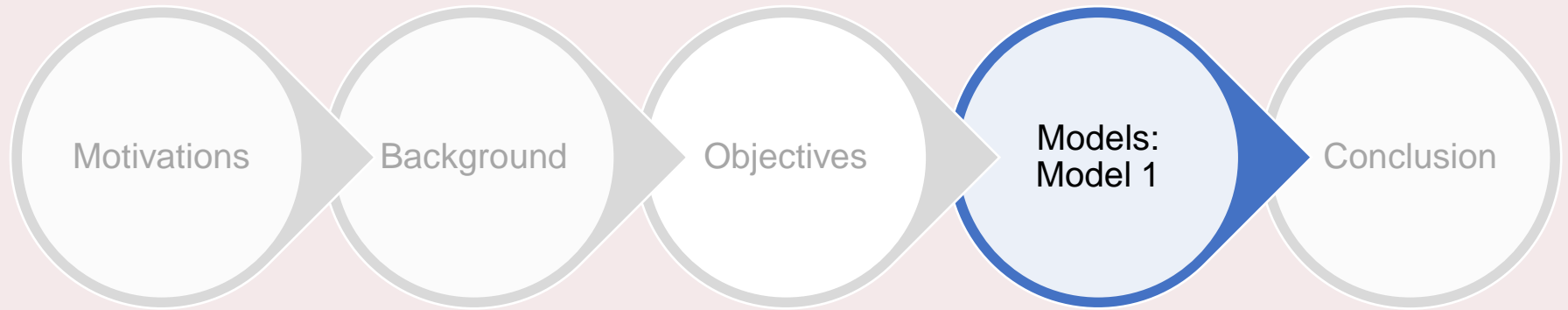
Quantify CD19⁺ B cell reconstitution in children post-HSCT, scaling for age-related effects and estimating time delay to B cell production

2

Identify the pharmacodynamics (PD) of rituximab in children with EBV post-HSCT

3

Quantify viral dynamics of EBV reactivation post-HSCT

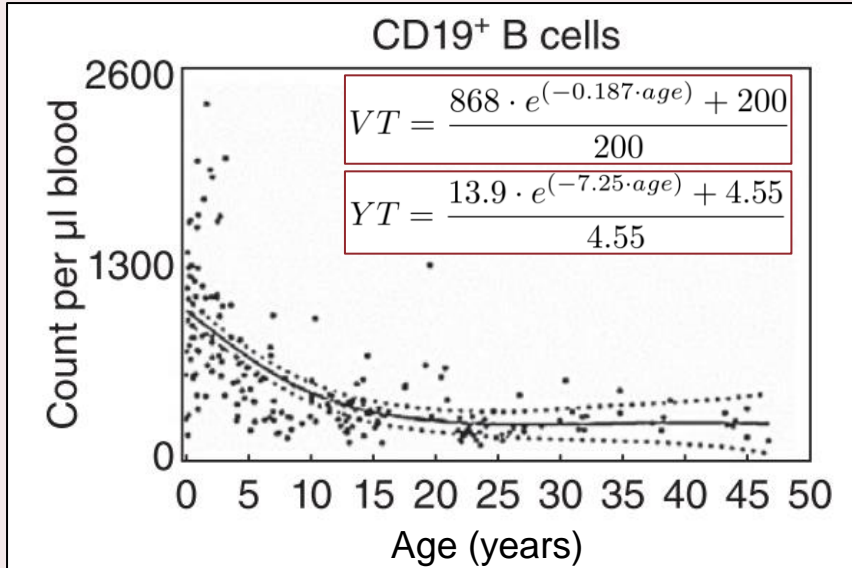


CD19⁺ B cell reconstitution in children post-HSCT



a) Scale for age-related effects

Developed B cell maturation function using non-linear least squares (VT: B cells, YT: Ki67)



Morbach et al (2010), *Clin & Exp Immuno.*[7]

b) Estimate time delay between HSCT and CD19⁺ B cell production

Tested two functions

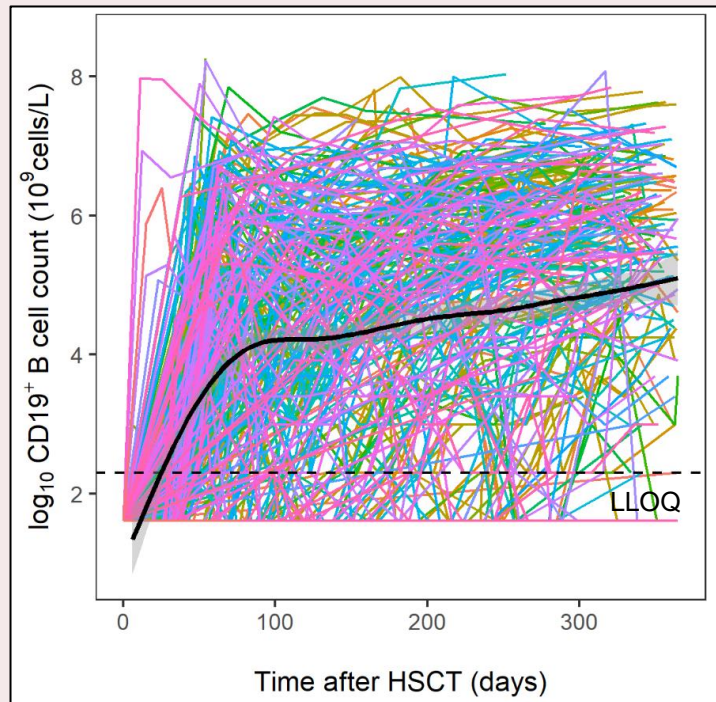
Equation	Parameters	Description
$y = \frac{1 - \exp\left(\frac{-2t}{\lambda_h}\right)}{1 + \exp\left(\lambda_r\left(1 - \frac{t}{\lambda_h}\right)\right)}$	λ_h (days) λ_r	Time to recovery of bone marrow output of CD19 ⁺ B cells Rate of recovery in bone marrow output of CD19 ⁺ B cells
$y = \frac{t^{Hill}}{t^{Hill} + T50^{Hill}}$	T50 (days) Hill co-efficient	Time to half-maximal bone marrow output of CD19 ⁺ B cells Steepness of slope of CD19 ⁺ B cell recovery

Hoare et al (2017), *Clin. Pharmacol. Ther* [8]

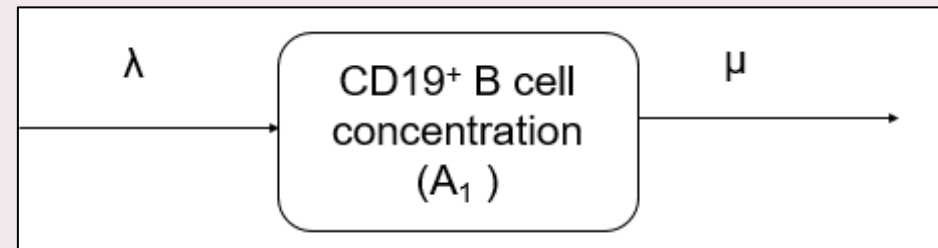
CD19⁺ B cell reconstitution in children post-HSCT

Data and Model

4115 CD19⁺ B cell counts from 359 children



Schematic and ODE



$$\frac{dA_1}{dt} = \lambda \cdot \left(\frac{T^\gamma}{T^\gamma + T50^\gamma} \right) - \mu(A_1)$$

$$\lambda = \lambda_{adult} \cdot VT \cdot YT$$
$$\mu = \mu_{adult} \cdot YT$$

CD19⁺ B cell reconstitution in children post-HSCT



Results

Description	Parameter/ Covariate (units)	Estimate/Effect size (% RSE)	% BSV (% RSE)	Bootstrap 95% CI	Bootstrap median
Number of CD19 ⁺ B cells at steady state	*Setpoint (x10 ⁶ cells/L)	*112	-	-	-
CD19 ⁺ B cell production rate constant	λ (x 10 ⁶ cells/day)	1.68 (2.45)	115.76 (0.0011)	1.36 – 2.05	1.68
CD19 ⁺ B cell death rate constant	μ (cells/day)	0.015 (3.00)	113.58 (0.0061)	0.013 – 0.018	0.015
Steepness of slope of CD19 ⁺ B cell recovery	Hill	4.17 (0.49)	-	4.05 – 4.29	4.17
Time to half-maximal output of CD19 ⁺ B cells from bone marrow	T50 (days)	58.9 (1.07)	137.48 (0.0033)	44.0 – 64.0	58.3
Effect of MAC on T50	T50_MAC	0.166 (10.12)	-	0.039 – 0.322	0.166
Effect of PID on T50	T50_PID	-0.551 (0.16)	-	-0.595 – to -0.457	-0.551
Effect of matched donor on T50	T50_Donor	-0.00998 (25.75)	-	-0.116 – 0.0786	0.00710

→ Higher than reported in adults
(Macallan et al (2005) Blood [9])

→ Aligns with experimental data
(Marie-Cardine et al (2008),
Clin. Imm. Ther [10])

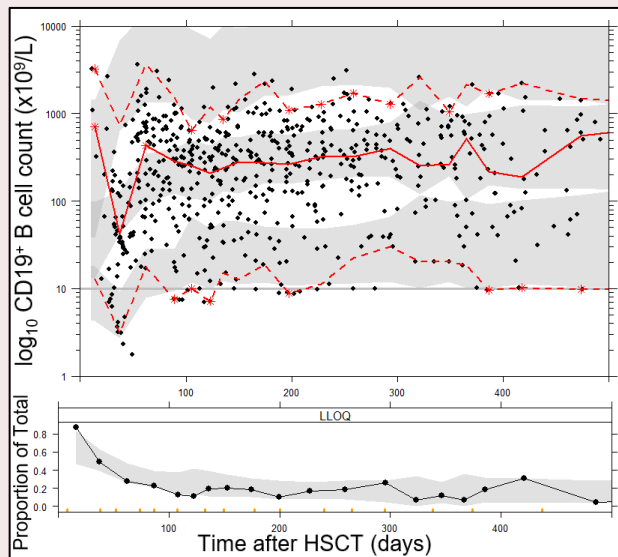
- *Denotes derived parameter (λ/μ).
- Initial condition of B cell compartment set to 5×10^6 cells/mL.
- Linearly parameterised covariate effects, i.e. multiplication of typical parameter value by (1 + effect size).

CD19⁺ B cell reconstitution in children post-HSCT

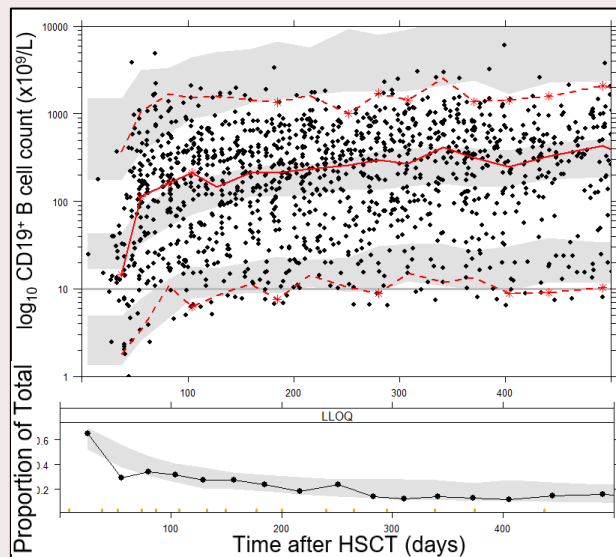


Model Evaluation by age group

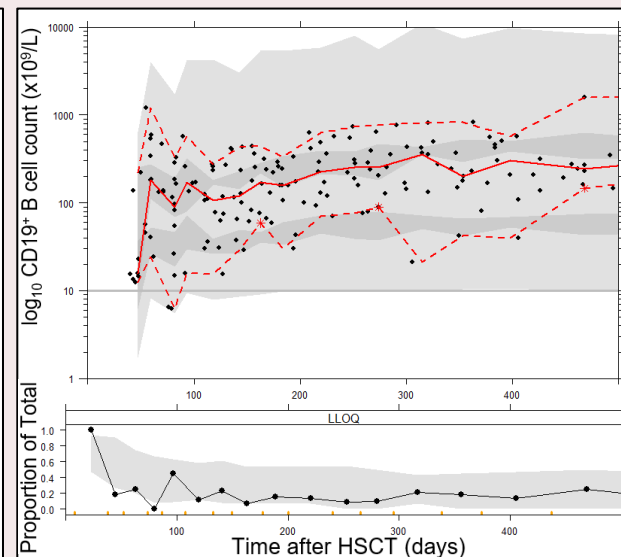
(A) Infants (< 2 years)



(B) Children (≥ 2 years and < 12 years)



(C) Adolescents (≥ 12 years)



Black dots are observed data, solid red line is observed median, dotted red line is observed 2.5th, 50th and 97.5th percentiles and grey shaded area is 95% prediction intervals for 2.5th, 50th and 97.5th percentiles.

1

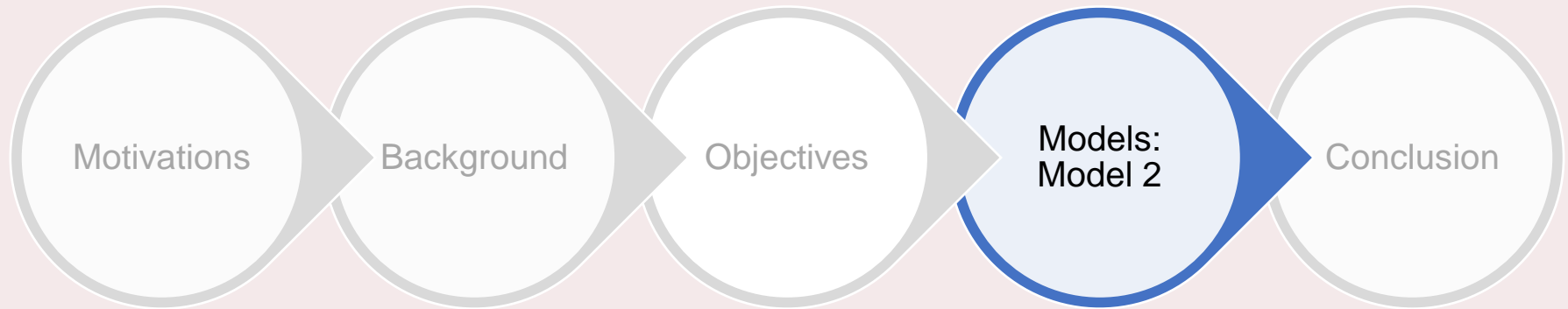
Built PD model to quantify CD19⁺ B cell reconstitution in children post-HSCT

2

Developed B cell maturation function based on CD19⁺ B cell count and Ki67 for *a priori* scaling of age-related effects

3

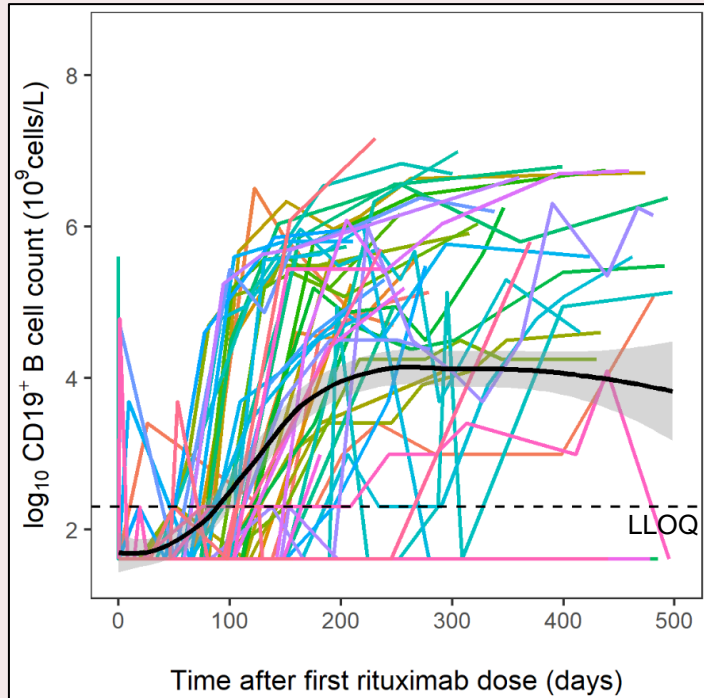
Incorporated Hill-type equation to estimate time delay between HSCT and CD19⁺ B cell production by the bone marrow



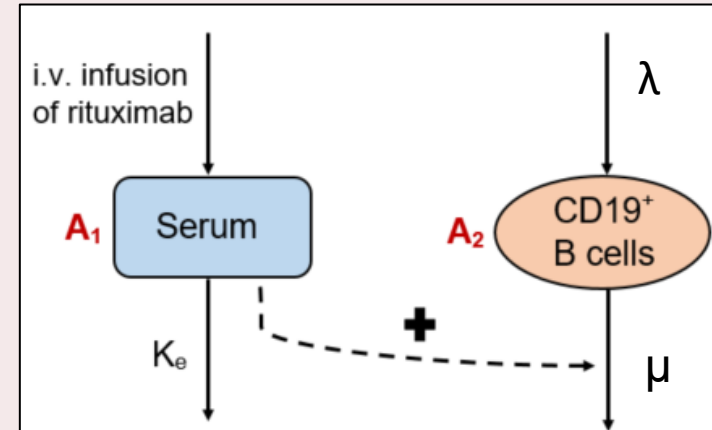
Rituximab PD in children with EBV post-HSCT

Data and Model

619 CD19⁺ B cell counts from 55 children



Schematic and ODE



$$\frac{dA_1}{dt} = -k_e \cdot A_1$$

$$\frac{dA_2}{dt} = \lambda \cdot \left(\frac{T^\gamma}{T^\gamma + T50^\gamma} \right) - \mu \cdot \left(1 + \frac{E_{max} \cdot A_1}{ED_{50} + A_1} \right) \cdot A_2$$

Rituximab PD in children with EBV post-HSCT



Results and Model Evaluation

Description	Parameter/ Covariate (units)	Estimate/Effect size
Number of CD19 ⁺ B cells at steady state	*Setpoint (x10 ⁶ cells/L)	*79.5
CD19 ⁺ B cell production rate constant	λ (x 10 ⁶ cells/day)	1.4
CD19 ⁺ B cell death rate constant	μ (cells/day)	0.017
Steepness of slope of CD19 ⁺ B cell recovery	Hill	3.18
Time to half-maximal output of CD19 ⁺ B cells from bone marrow	T50 (days)	44.8
Rituximab elimination rate constant	K _e (/day)	0.109
Maximum killing effect of rituximab on CD19 ⁺ B cells	E _{max}	84.4
Rituximab dose producing 50% of maximum killing effect	ED50 (mg)	0.921
Effect of MAC on T50	T50_MAC	0.954
Effect of PID on T50	T50_PID	-0.474
Effect of matched donor on T50	T50_Donor	-0.465

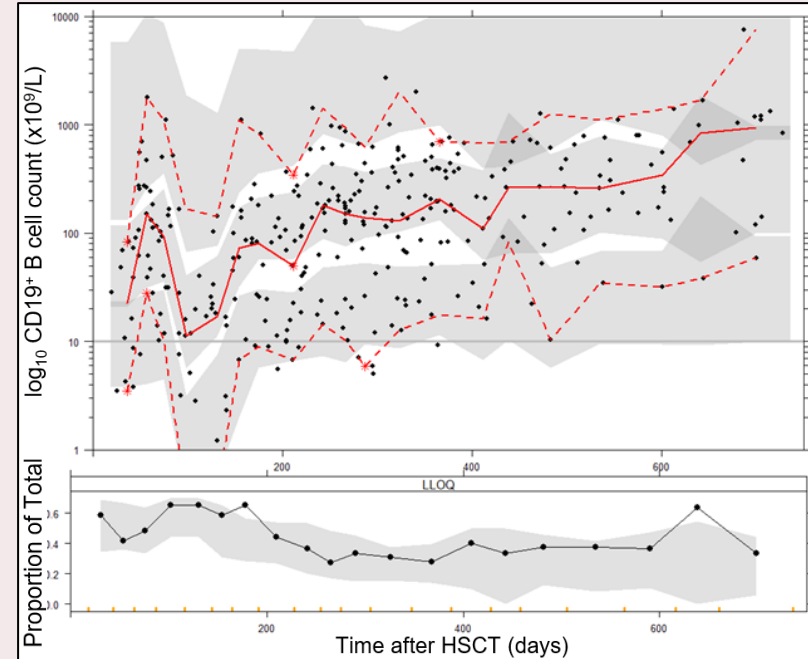
Estimated half-life (children):

-26±3 days

-19.3 days (11.1–73.7)

-6.35 days Current study

Higher in non-rituximab patients



1

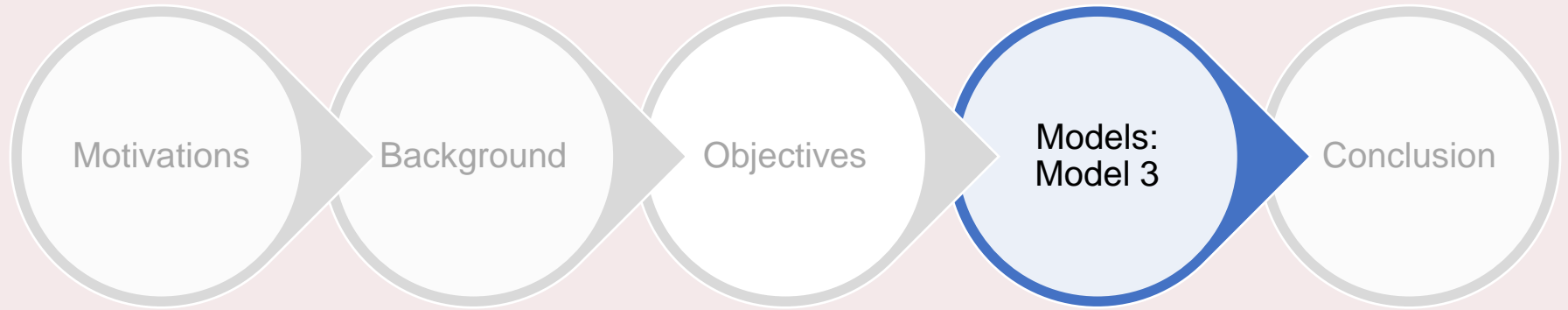
Built K-PD model to quantify rituximab PD in children with EBV post-HSCT

2

Incorporated rituximab drug effect using E_{\max} model

3

Estimated elimination half-life of 6.35 days for rituximab



Dynamics of EBV reactivation post-HSCT

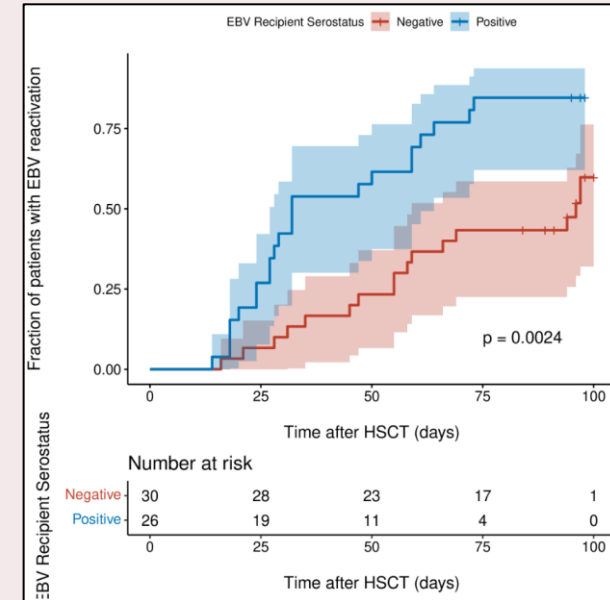
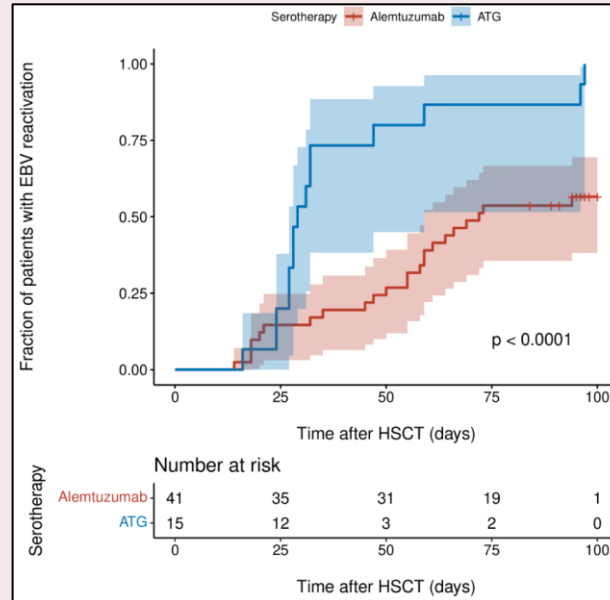
Cox proportional hazards (PH): Cox-PH model for EBV reactivation in first 100 days post-HSCT in 56 children

Covariates tested:

- HSCT**
- Donor type
 - HSC source
 - Cond'g regimen type

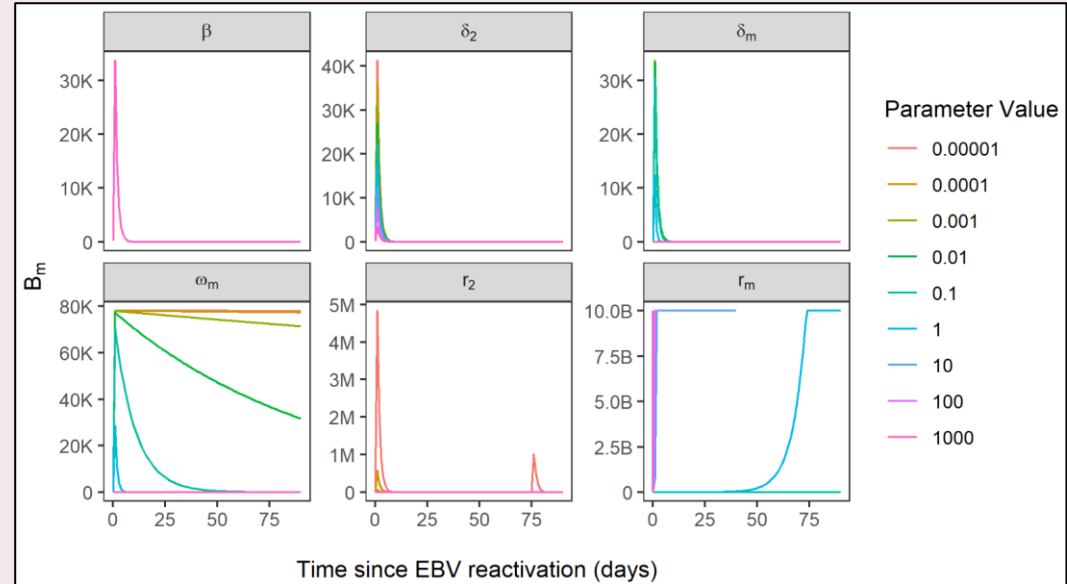
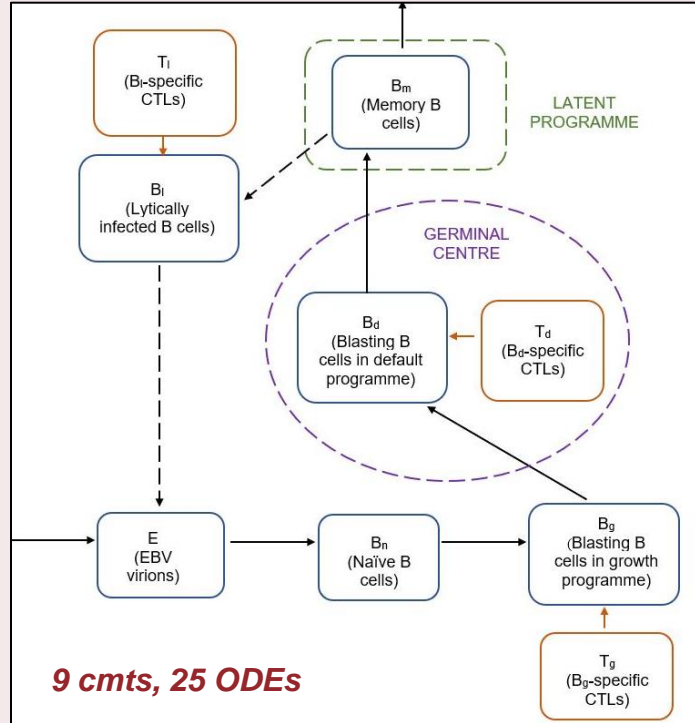
- Patient**
- PID diagnosis
 - Age
 - Donor/Recipient EBV serostatus

- Immune cells + drug admin.**
- AUC_{0-100} for ALC/ CD19⁺ B/ CD4⁺ T/CD8⁺ T cells
 - No. of Rituximab Doses
 - Pre-HSCT Alemtuzumab or Anti-thymocyte globulin (ATG)



Dynamics of EBV reactivation post-HSCT

Sensitivity analysis: Previously reported **mechanistic mathematical model** of EBV kinetics



β , NK effect on EBV inf'n; δ_2 , CTL killing rate of infected B cells; δ_m , death rate of inf'd memory B cells; ω_m , memory B cell reactivation rate; r_2 , CTL activation rate against infected B cells; r_m , memory B cell prolif'n rate

Dynamics of EBV reactivation post-HSCT

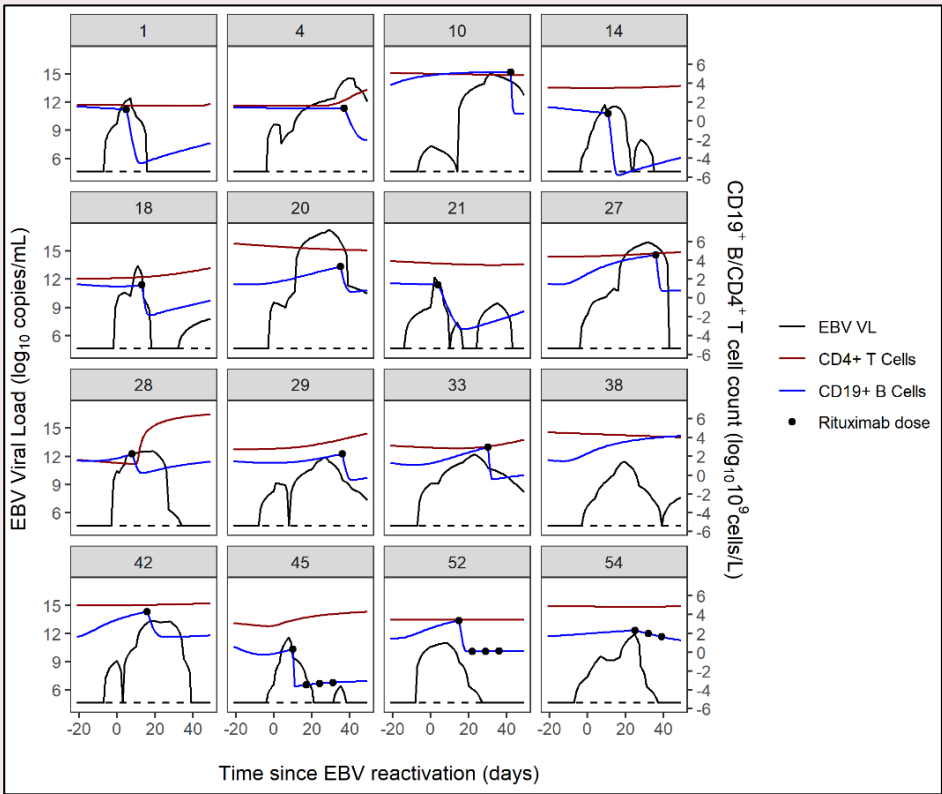


Pharmacometric approach

Rituximab PD model + knowledge from sensitivity analysis of previously reported model



Pharmacometric model of EBV reactivation dynamics post-HSCT



Dynamics of EBV reactivation post-HSCT

ODEs and Individuals Fits

$$\frac{dA_1}{dt} = -k_e \cdot A_1$$

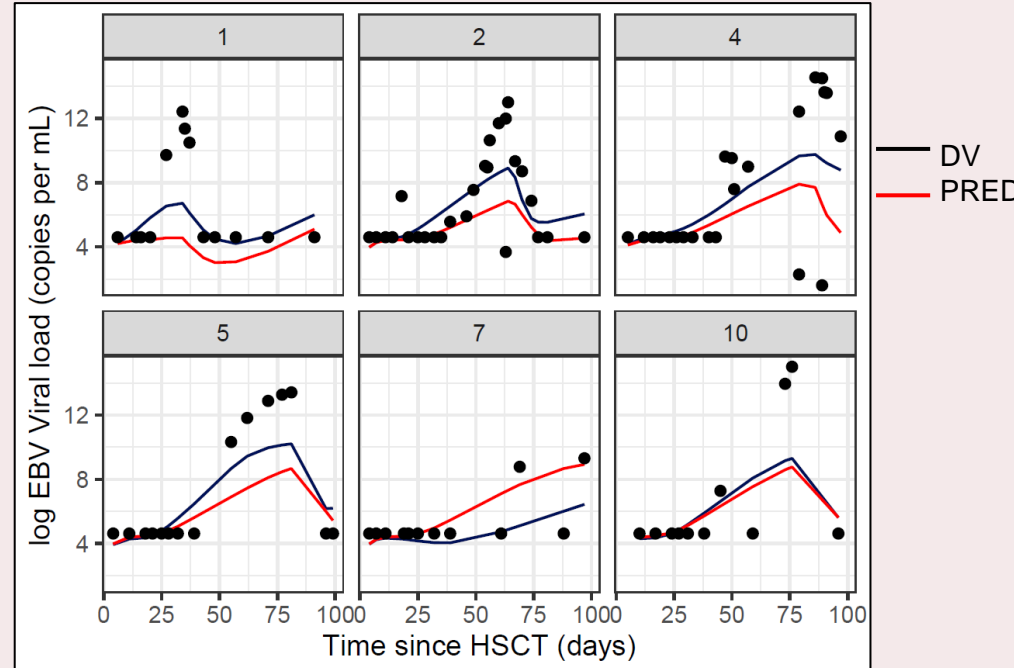
$$\frac{dA_2}{dt} = \lambda \cdot \left(\frac{T^\gamma}{T^\gamma + T^{50\gamma}} \right) - \mu \cdot \left(1 + \frac{E_{max} \cdot A_1}{ED_{50} + A_1} \right) \cdot A_2$$

$$\frac{dA_3}{dt} = K \cdot A_2 \cdot -\delta \cdot \left(1 + \frac{E_{max} \cdot A_1}{ED_{50} + A_1} \right) \cdot A_3 - Q \cdot A_3 \cdot A_5$$

$$\frac{dA_4}{dt} = N \cdot \delta \cdot A_3 - C \cdot A_4$$

$$\frac{dA_5}{dt} = R \cdot A_3 - D \cdot A_5$$

K , Production rate of infected B cells; δ , Death rate of infected B cells;
 Q , CTL killing rate of infected B cells; N , viral burst size; C , viral clearance rate;
 R , Proliferation rate of EBV-specific CTLs ; D , CTL death rate



1

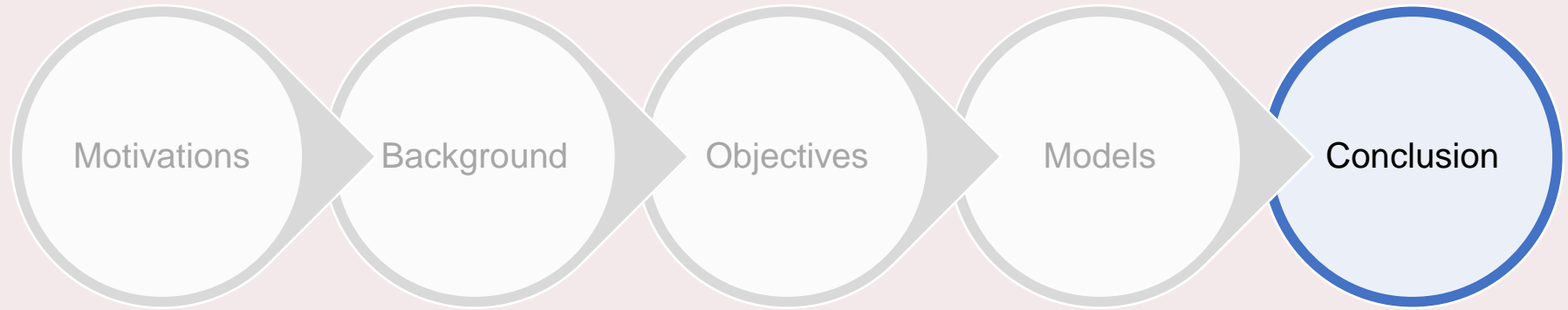
Pre-HSCT ATG and recipient EBV seropositivity are significant risk factors for EBV reactivation in first 100 days post-HSCT in Cox-PH model

2

Parameters related to memory B cells and CTLs drive EBV viral load according to sensitivity analysis

3

Developing pharmacometric model to quantify dynamics of EBV reactivation post-HSCT



Unmet Need



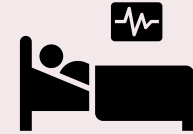
- B cells
- Children
- Pharmacometrics

Our Work

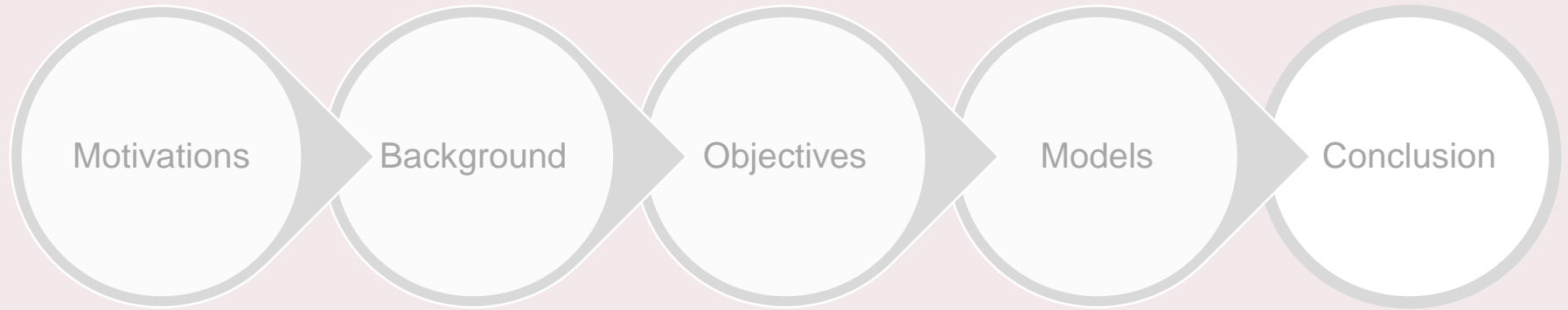


- Built **pharmacometric** models with **age scaling** for:
 - CD19⁺ **B cell** reconstitution in **children** post-HSCT
 - **Rituximab** PD in **children** with EBV post-HSCT
 - **EBV reactivation** in **children** post-HSCT
- **Future work** on bioequivalence in **rituximab biosimilars**

Clinical Impact



- **PID diagnosis, MAC and matched donor** affect B cell recovery
- Inform **rituximab dosing**
- Models can **predict** patients' B cell recovery **trajectories**



1. Rosser EC, Wilkinson MG. B cells as a therapeutic target in paediatric rheumatic disease. *Frontiers in immunology*. 2019;10:214.
2. Odumade OA, Hogquist KA, Balfour HH. Progress and problems in understanding and managing primary Epstein-Barr virus infections. *Clinical microbiology reviews*. 2011 Jan 1;24(1):193-209.
3. Gibiansky E, Gibiansky L, Chavanne C, Frey N, Jamois C. Population pharmacokinetic and exposure–response analyses of intravenous and subcutaneous rituximab in patients with chronic lymphocytic leukemia. *CPT: pharmacometrics & systems pharmacology*. 2021 Aug;10(8):914-27.
4. Ng CM, Bruno R, Combs D, Davies B. Population pharmacokinetics of rituximab (anti-CD20 monoclonal antibody) in rheumatoid arthritis patients during a phase II clinical trial. *The Journal of Clinical Pharmacology*. 2005 Jul;45(7):792-801.
5. Barth MJ, Goldman S, Smith L, Perkins S, Shiramizu B, Gross TG, Harrison L, Sanger W, Geyer MB, Giulino-Roth L, Cairo MS. Rituximab pharmacokinetics in children and adolescents with de novo intermediate and advanced mature B-cell lymphoma/leukaemia: a children's oncology group report. *British journal of haematology*. 2013 Sep;162(5):678-83.
6. Pan S, Yu H, Surti A, Cheng I, Marks SD, Brogan PA, Eleftheriou D, Standing JF. Pharmacodynamics of rituximab on B lymphocytes in paediatric patients with autoimmune diseases. *British journal of clinical pharmacology*. 2019 Aug;85(8):1790-7.
7. Morbach H, Eichhorn EM, Liese JG, Girschick HJ. Reference values for B cell subpopulations from infancy to adulthood. *Clinical & Experimental Immunology*. 2010 Nov;162(2):271-9.
8. Hoare RL, Veys P, Klein N, Callard R, Standing JF. Predicting CD4 T-cell reconstitution following pediatric hematopoietic stem cell transplantation. *Clinical Pharmacology & Therapeutics*. 2017 Aug;102(2):349-57.
9. Macallan DC, Wallace DL, Zhang Y, Ghattas H, Asquith B, de Lara C, Worth A, Panayiotakopoulos G, Griffin GE, Tough DF, Beverley PC. B-cell kinetics in humans: rapid turnover of peripheral blood memory cells. *Blood*. 2005 May 1;105(9):3633-40.
10. Marie-Cardine A, Divay F, Dutot I, Green A, Perdrix A, Boyer O, Contentin N, Tilly H, Tron F, Vannier JP, Jacquot S. Transitional B cells in humans: characterization and insight from B lymphocyte reconstitution after hematopoietic stem cell transplantation. *Clinical immunology*. 2008 Apr 1;127(1):14-25.
11. Kania SP, Silva JMF, Charles OJ, Booth J, Cheung SYA, Yates JWT, Worth A, Breuer J, Klein N, Amrolia PJ, Veys P, Standing JF. Epstein-Barr Virus Reactivation After Paediatric Haematopoietic Stem Cell Transplantation: Risk Factors and Sensitivity Analysis of Mathematical Model. *Frontiers in Immunology*. 2022. doi: 10.3389/fimmu.2022.903063
12. Akinwumi, S (2018). Modelling the Kinetics of EBV in Primary Carriers and Transplant Recipients. PhD thesis. Department of Mathematical and Statistical Sciences, University of Alberta
13. Wang Y, Zhou Y, Brauer F, Heffernan JM. Viral dynamics model with CTL immune response incorporating antiretroviral therapy. *Journal of mathematical biology*. 2013 Oct;67(4):901-34.

Acknowledgements

PhD Supervisors

Prof Joe Standing

Prof Nigel Klein



Dr James Yates

Dr Amy Cheung



Clinical collaborators

GOSH BMT Unit

Juliana Silva, Reem Elfeky,

Persis Amrolia, Paul Veys



Addenbrookes Hospital

Amos Burke



GOSH DRIVE

John Booth

Mohsin Shah



London Pharmacometrics Interest Group

Funding

MRC iCASE studentship (MR/R015759/1)

AstraZeneca



Patients and their families

